



Acute lymphoid leukemia etiopathogenesis

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Abstract

Acute lymphoid leukemia (ALL) is a type of hematological neoplasm that affects the precursor cells of strains B, T and NK, with a higher incidence in the pediatric range. The pathophysiology of ALL is characterized by chromosomal abnormalities and genetic alterations involved in the differentiation and proliferation of lymphoid precursor cells. Despite the lack of information in the literature, it is believed that leukemogenesis originates from a complex interaction between environmental and genetic factors, which combined lead to cellular modifications. Environmental factors have been evaluated as possible predisposing factors in the development of ALL but there are still conflicting results in the world literature. In this context, the aim of the present review is to discuss the major exogenous factors regarding ALL.

Keywords Leukemia · Childhood · Infection · Environment

Introduction

Childhood and juvenile cancer, which affects individuals between 0 and 19 years old, consists of a set of diseases that have their own characteristics in relation to the cells that make up the tumors (histological type) and clinical behavior of the disease [1]. In most populations, childhood and juvenile cancer accounts for 1% to 4% of all malignant tumors and in developing countries, where the child population reaches 50%, this proportion of childhood cancer accounts for 3% to 10% of all cancers. Worldwide, among childhood cancer types, leukemia is common in most populations, corresponding to 25–35% of cases [2].

Leukemias are a group of different hematologic diseases with a different biological concept, clinical presentation, prognosis and treatment response, characterized by the

presence of an abnormal cell population suppressing the normal production of cellular components of the hematopoietic system [3].

Leukemias originate from hematopoietic stem cells (HSCs) and precursors in the bone marrow (BM), that promote leukemic cell proliferation and infiltration [4, 5]. In general, hematological malignancies are classified according to lineage, degree of maturation and form of cellular involvement in the BM. Myeloid lineage neoplasms may include granulocytes (neutrophils, eosinophils, basophils), monocytes, erythrocytes, platelets (megakaryocyte derivatives) and mast cells. In contrast, the lymphoid lineage corresponds to B, T and natural killer (NK) cells [6].

Acute lymphoblastic leukemia (ALL) is the most common childhood malignancy [7]. The majority of ALL occurs in healthy individuals and predisposing factors, such as hereditary genetic susceptibility or environmental exposure, have been identified in only a few patients. It is characterized by chromosomal abnormalities and genetic changes involved in lymphoid differentiation and proliferation precursor cells [8].

ALL and lymphoblastic lymphoma (LBL) are biologically comparable and treated the same [9], ALL/LBL comprises B-lineage, T-lineage, and uncommon variants (i.e., NK-lineage, early T progenitor ALL [a provisional diagnostic category]) [10].

The ALL pathophysiology involves complex environmental as well as genetic mechanisms at different scales, and

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there must also be a complex relationship between these factors [11]. Along with response to treatment, these abnormalities are important prognostic factors [8].

Epidemiologic studies of acute leukemias in children have examined possible risk factors, including genetic, infectious and environmental, in an attempt to determine their etiology [12]. It is believed that ALL may originate from interactions between exogenous factors. Therefore, this review proposes to clarify the understanding of ALL regarding environmental factors.

Acute lymphoid leukemia

ALL is a malignant disorder that originates from a single haematopoietic precursor affecting the B or T cell line. These cells can acquire a series of genetic alterations that may disrupt normal maturation processes, leading to the blockade of differentiation and transformed cell proliferation [13].

The distribution of ALL categories is B lineage (85%), T lineage (10–15%) and NK lineage (<1%). The incidence of this disease varies worldwide but may be influenced, in part, by diagnostic and reporting differences [14]. The incidence of ALL is higher in Hispanic (43 cases/million) and White (Caucasian) Americans (36 cases/million) than in Black Americans (15 cases/million) and Asians [15]. The peak incidence of ALL occurs between 2 and 5 years of age and it is more common among boys than girls [15, 16].

In the United States, approximately 6000 new ALL cases have been estimated, at a ratio of 1.3 male to 1 female [17]. Most patients are children and 60% cases occur in individuals under 20 years old [7, 18, 19].

ALL, like cancer in general, probably originates from interactions between exogenous and endogenous exposure and genetic susceptibility [20]. ALL pathogenesis occurs from important genetic lesions in genes involved in T or B lymphocyte differentiation [7]. The challenge is to identify exposure elements and relevant inherited genetic variants and also to decipher how and when these factors contribute to the multi-stage natural history of ALL from initiation (usually in the womb) to disease manifestation [21].

The classification for ALL was proposed by the World Health Organization (WHO) (1997) in an attempt to account for the morphology and cytogenetic profile of the leukemic blasts and identified three types of ALL: B lymphoblastic, T lymphoblastic and Burkitt-cell Leukemia [22]. In 2008, Burkitt-cell leukemia was eliminated, and became only Burkitt's lymphoma and B-lymphoblastic leukemia was again divided into two subtypes: B-ALL with recurrent genetic abnormalities and B-ALL without further specification. [6]. In 2016, two new provisional entities were added to the list of recurrent genetic abnormalities and the hypodiploid was

redefined as either low hypodiploid or hypodiploid with *TP53* mutations [23].

Pathophysiology of ALL

Leukemia and other cancers share the same biological characteristic which is clonality. Molecular changes are necessary for the development of cancer and they can alter the components of the signaling pathway, that lead to the emission of proliferative signals, even when no more cells are needed, inappropriately activating cell growth, DNA replication and cell division [11].

The development of a malignant hematological disease probably involves a mutation in a critical gene of cell proliferation, differentiation and/or survival in a hematopoietic progenitor [24]. Most leukemia mutations are acquired and occur in a lymphoid cell progenitor; mutated genes are less often inherited (1–5% of leukemias) and this involves a numerical chromosomal abnormality, such as 21 trisomy [25].

When an oncogene is activated by mutation, the encoded protein is structurally modified and generally shows increased transformative activity, remaining in its active state, continuously transmitting its signals through the interaction of tyrosines and/or threonine kinases. These signals induce incessantly continued cell proliferation [11].

There are mutations that suppress gene function and occur in tumor suppressor genes, such as *TP53*. However, less than 3% of childhood patients and an average of 8% of adult ALL patients have mutations in *TP53* [26].

Numeric chromosomal abnormalities as well as structural rearrangements (translocations) commonly occur in ALL. Significant cytogenetic abnormalities in B cell precursors are associated with poor ALL prognosis, including t(9; 22) or Philadelphia chromosome, with frequency directly proportional to age [27]; t(4; 11), related to the mixed lineage leukemia (MLL) gene, common in childhood and also associated with myeloid leukemia [28, 29]; hypodiploidy [30]; and chromosome 8 trisomy in adults diagnosed with ALL [31].

In addition, the genetic alterations associated with ALL are mainly localized at sites where oncogenes exist, for example: the MLL oncogene MLL related to ALL in children [32]; t(8; 14) translocation associated with gene dysregulation of the C-MYC oncogene [33]; *TP53* tumor suppressor gene mutations [34]; and deletions and inversions, such as the transcription factor *PAX5* deletions, present in at least 30% of B cell precursor ALL [35].

Aberrant methylation of CpG islands in gene promoter regions has been identified in ALL cell lines and is considered important since methylation of CpG dinucleotides near transcription initiation sites may silence gene expression

[36]. Thus, hypermethylation of tumor suppressor genes and hypomethylation of oncogenes can trigger leukemias.

Another important mechanisms of ALL development are the modification of angiogenesis [37], signal transduction in interaction with tyrosine kinase receptors and apoptosis regulatory molecules [38], as is the case of the *BCL2* gene, which encodes a cytoplasmic protein located in the mitochondria and increases cell survival by inhibiting apoptosis.

There are cases of complications from long-term treatment observed in cancer patients, which is the increased risk of developing hematological malignancies [39]. They usually manifest as acute leukemias or myelodysplastic syndromes and are frequently high, possibly due to increased use of genotoxic agents in antitumor therapies and increased survival in other cancers [40].

Increasing evidence supports a multi-step process in leukemogenesis, with sequential steps and a series of changes in oncogenes, tumor suppressor genes and microRNA genes in tumor cells [24, 41]. Unlike genes involved in cancer development, genes for microRNAs do not encode proteins; their products are small RNA molecules (single stranded 21–23 nucleotides) that recognize and bind to messenger RNA (mRNA) nucleotide sequences, blocking protein translation and thereby regulating gene expression [41, 42].

Several microRNAs have been implicated in ALL pathogenesis [43–45]. In this context, Li, Li [46] identified an increase in miR-708, miR-210 and miR-181b microRNA expression in B-line precursor ALL cells (common ALL). Moreover, they demonstrated that miR-708 expression is related to the high-risk ALL group, when compared to low-risk groups, by regulating the expression of the ciliary neurotropic factor receptor (*CNFTR*), neuronatin (*NNAT*) and guanine nucleotide-binding protein subunit gamma (*GNG12*) genes.

Molecular changes required for leukemia development are rare phenomena when considering the large number of target cells susceptible to genetic modification [24]. It is important to mention that when referring to the origin of cancer, in the case of ALL, two terms must be referenced: the original cell and the leukemic stem cell.

Environmental factors

Although little is known about the etiology of ALL, the multifactorial behavior of the disease suggests that risk factors contribute to its development, such as environmental and/or genetic risk factors [47]. On the other hand, there are three highlighted hypotheses: population mix [48], late infection [49] and hygienic-sanitary [50], suggesting the involvement of the immune system in the ALL etiology.

Studies have reported an increased incidence of ALL in association with advanced paternal age and maternal

fetal loss (hazard ratios ≤ 1.1 for either) [11–13]. Studies of the relationship between childhood leukemia and urban/rural status, population density and other possible etiologic factors (eg, environmental exposures, abnormal immune response to common infections) have yielded inconsistent results [50–56].

In most cases, childhood ALL is not considered to be a familial disease [57]. Importantly, because ALL is a disease with a low incidence in the general population, even a large increase in the relative risk among siblings does not translate into a high likelihood of developing leukemia. Certain genetic syndromes (eg, Down syndrome, neurofibromatosis type 1, Bloom syndrome, ataxia-telangiectasia) [58, 59], rare germline mutations in *PAX5*, *ETV6*, and *TP53* and polymorphic variants of *ARD5B*, *CDKN2A* and *IKZF1* (the gene encoding Ikaros) are associated with an increased risk of leukemia [60–64].

One of the most related environmental factors associated with an increased risk for ALL is the involvement of ionizing radiation, which is more prevalent in pediatric leukemias, especially in ALL and acute myeloid leukemia (AML) [12, 65]. Potential exposure of children to ionizing radiation may occur during the gestational phase or in the postnatal period [12]. Other risk factors already accepted include prenatal x-ray exposure and postnatal exposure to high doses of radiation [66].

In a study associating haplotypes with patients that were exposed to radiation, Chokkalingam, Bartley [67] observed that, when analyzing 32 genes responsible for cell cycle repair pathways, 4 haplotypes from *APEX1*, *BRCA2*, *RAD51* and *ERCC2* genes demonstrated a risk association. They also showed that 3 genes (*NBN*, *XRCC4* and *CDKN2A*) were associated with structural and numerical genetic alterations in ALL patients, showing genetic susceptibility regarding ionizing radiation.

The hypotheses of population mixture and late infection suggest that a poor immune system in the early stages of human development may cause abnormal immune responses against infections, allowing an altered cell to develop. Both hypotheses are similar to the sanitary hygienic hypothesis, which explains the emergence of the original cell due to increased allergy frequency during the first years of life. Many studies support the hypothesis of infections and the immune system as ALL etiological factors [47, 50, 54, 68, 69]. However, little is known about the role of genes in this etiology.

Although there are studies with conflicting results, environmental factors have been investigated as predisposing to the development of ALL. Moreover, adult leukemia risk factors differ from those of children. For adult-onset leukemia, the most established environmental risk factor is exposure to ionizing radiation. Benzene, agricultural exposures, smoking and the consumption of alcohol, cigarettes

and illicit drugs during pregnancy have been described as predisposing factors for childhood ALL [70–75].

A common type of DNA change that can lead to leukemia is known as a chromosome translocation, for example, a translocation seen in some cases of childhood ALL is a swap of DNA between chromosomes 9 and 22, which leads to what is known as the Philadelphia [76], and the review by Quiroz, Aldoss [77] describes chromosomal abnormalities and specific genomic landscape in Latinos with ALL and their association with unfavorable prognosis, focusing on Philadelphia chromosome-positive (Ph⁺) ALL. These authors compared ALL distribution throughout the various countries in Latin America in an attempt to shed some epidemiological light on the genetic ancestry of ALL.

Growing evidence from epidemiological studies suggests that the increased leukemia rate is probably related to an abnormal immune response to infections early in life and viruses have been suggested to play a role in the ALL pathogenesis [49, 78]. Some biological agents, such as viruses, as the Epstein Barr virus (EBV) and the human T cell lymphotropic virus 1 (HTLV-1), also remain as risk factors for ALL. The data suggest that a dysfunctional immune response to a common childhood pathogen(s), such as bacterial or viral infection, is a final step leading to leukemia [20, 50].

Many epidemiological studies have attempted to identify risk factors in the etiopathogenesis of ALL. Belson, Kingsley [12] presented an elegant review that discusses the environmental risk factors including some of the most researched and also controversial topics such as hydrocarbons and pesticides, alcohol, ionizing and non-ionizing radiation, smoking and use of illicit drugs. In addition, they also reviewed genetic and infectious risk factors and others. Studies of environmental exposures and childhood cancer do not depend only on the records of the maternal address at birth or at the time of diagnosis, but on whether there is residential mobility to assess exposures in early childhood [79, 80].

The relationship between the immune system and ALL is a complex process that involves the interaction of many components such as epithelial barriers, cells, proteins, cytokines and variations in the genes of these cells can affect the development and function of immune responses, and thus increase susceptibility to ALL [68, 81].

There is evidence that advanced parental age is associated with increased childhood ALL risk and this association was most marked among children aged 1–5 years. Employing datasets with cytogenetic information may further elucidate the involvement of each parental component and clarify underlying mechanisms [82].

A study by Orsi et al. [83] included 7847 ALL cases and 11,667 controls aged 1–14 years old and investigated the associations between childhood ALL) and several factors related to early stimulation of the immune system, that is,

farm residence and regular contacts with farm animals (livestock, poultry) or pets in early childhood. However, there was no evidence of significant association with agricultural residence and regular contact with animals in the first year.

In this context, there is a concern in the developing countries, with a broad heterogeneous territorial population exposed to different infectious agents and hygiene levels, and also a concern with children, that, in addition to being exposed to common infectious diseases in this age group, are prone to the development of regional diseases, such as parasitic diseases, mainly due to climatic and sanitation differences.

Although the recognition of leukemia among childhood diseases is well established, as well as the role of treatment groups for this disease, very little is known about the involvement of the environment in the development of the disease. As pointed out in this review, the true biological profile of childhood leukemia is unknown and in-depth studies should be made regarding different geographical regions, exposure to risk factors, such as exposure to ionizing rays, xenobiotics and chemicals, in addition to the presence of infectious microorganisms.

References

1. Little J (1999) Epidemiology of childhood cancer. IARC Scientific Publications, Lyon
2. Howlader N et al SEER cancer statistics review, 1975-2011, vol 2014. National Cancer Institute, Bethesda
3. Polychronakis I et al (2013) Work-related leukemia: a systematic review. *J Occup Med Toxicol* 8(1):14
4. Konopleva MY, Jordan CT (2011) Leukemia stem cells and micro-environment: biology and therapeutic targeting. *J Clin Oncol* 29(5):591–599
5. Azizidoost S et al (2014) Bone marrow neoplastic niche in leukemia. *Hematology* 19(4):232–238
6. Vardiman JW et al (2009) The 2008 revision of the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia: rationale and important changes. *Blood* 114(5):937–951
7. Pui CH, Robison LL, Look AT (2008) Acute lymphoblastic leukaemia. *Lancet* 371(9617):1030–1043
8. Malard F, Mohty M (2020) Acute lymphoblastic leukaemia. *Lancet* 395(10230):1146–1162
9. McNeer JL, Bleyer A (2018) Acute lymphoblastic leukemia and lymphoblastic lymphoma in adolescents and young adults. *Pediatr Blood Cancer* 65(6):e26989
10. Campo EH, Jaffe ES, Pileri SA, Stein H, Thiele J, Arber D, Hasserjian R, Le Beau M (2017) WHO classification of tumours of haematopoietic and lymphoid tissues, 4th edn, Lyon
11. Gallegos-Arreola M, et al. (2013) Pathophysiology of acute lymphoblastic leukemia. In: Mejia-Arangue JM (ed) Clinical epidemiology of acute lymphoblastic leukemia – From the molecules to the clinic
12. Belson M, Kingsley B, Holmes A (2007) Risk factors for acute leukemia in children: a review. *Environ Health Perspect* 115(1):138–145

13. Graux C (2011) Biology of acute lymphoblastic leukemia (ALL): clinical and therapeutic relevance. *Transfus Apher Sci* 44(2):183–189
14. Steliarova-Foucher E, Colombet M, Ries LAG, Moreno F, Dolya A, Bray F, Hesselting P, Shin HY, Stiller CA (2017) IICC-3 contributors. International incidence of childhood cancer, 2001–10: a population-based registry study. *Lancet Oncol*. 18(6):719–731. [https://doi.org/10.1016/S1470-2045\(17\)30186-9](https://doi.org/10.1016/S1470-2045(17)30186-9)
15. Ries LG, Smith MA, Gurney JG et al (eds) (1999) Cancer incidence and survival among children and adolescents: United States SEER Program 1975–1995. National Cancer Institute, SEER Program. NIH Pub. No. 99-4649. Bethesda, MD
16. Institute, N.C., SEER Cancer Statistics Review, 1975-2013. 2015, Bethesda, MD
17. Siegel R, Naishadham D, Jemal A (2012) Cancer statistics, 2012. *CA Cancer J Clin* 62(1):10–29
18. Stanulla M, Schrappe M (2009) Treatment of childhood acute lymphoblastic leukemia. *Semin Hematol* 46(1):52–63
19. Hunger SP et al (2012) Improved survival for children and adolescents with acute lymphoblastic leukemia between 1990 and 2005: a report from the children's oncology group. *J Clin Oncol* 30(14):1663–1669
20. Inaba H, Greaves M, Mullighan CG (2013) Acute lymphoblastic leukaemia. *Lancet* 381(9881):1943–1955
21. Greaves MF, Wiemels J (2003) Origins of chromosome translocations in childhood leukaemia. *Nat Rev Cancer* 3(9):639–649
22. Harris NL et al (1999) The world health organization classification of neoplastic diseases of the hematopoietic and lymphoid tissues. Report of the clinical advisory committee meeting, Airlie House, Virginia, November, 1997. *Ann Oncol* 10(12):1419–1432
23. Arber DA et al (2016) The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood* 127(20):2391–2405
24. Greaves M (2002) Childhood leukaemia. *BMJ* 324(7332):283–287
25. Seewald L et al (2012) Acute leukemias in children with down syndrome. *Mol Genet Metab* 107(1–2):25–30
26. Chiaretti S, Zini G, Bassan R (2014) Diagnosis and subclassification of acute lymphoblastic leukemia. *Mediterr J Hematol Infect Dis* 6(1):e2014073
27. Rix U et al (2013) A target-disease network model of second-generation BCR-ABL inhibitor action in Ph+ ALL. *PLoS One* 8(10):e77155
28. Armstrong SA et al (2002) MLL translocations specify a distinct gene expression profile that distinguishes a unique leukemia. *Nat Genet* 30(1):41–47
29. Armstrong SA et al (2003) Inhibition of FLT3 in MLL. Validation of a therapeutic target identified by gene expression based classification. *Cancer Cell* 3(2):173–183
30. Nachman JB et al (2007) Outcome of treatment in children with hypodiploid acute lymphoblastic leukemia. *Blood* 110(4):1112–1115
31. Bakshi SR et al (2012) Trisomy 8 in leukemia: a GCRI experience. *Indian J Hum Genet* 18(1):106–108
32. Ayton PM, Cleary ML (2001) Molecular mechanisms of leukemogenesis mediated by MLL fusion proteins. *Oncogene* 20(40):5695–5707
33. Angi M, Kamath V, Yuvarani S, Meena J, Sitaram U, Manipadam MT, Nair S, Ganapule A, Fouzia NA, Abraham A, Viswabandya A, Poonkuzhali B, George B, Mathews V, Srivastava A, Srivastava VM (2017) The t(8;14)(q24.1;q32) and its variant translocations: a study of 34 cases. *Hematol/Oncol Stem Cell Therapy* 10(3):126–134. <https://doi.org/10.1016/j.hemonc.2017.03.002>
34. Vilas-Zornoza A et al (2011) Frequent and simultaneous epigenetic inactivation of TP53 pathway genes in acute lymphoblastic leukemia. *PLoS One* 6(2):e17012
35. Heltemes-Harris LM et al (2011) Ebf1 or Pax5 haploinsufficiency synergizes with STAT5 activation to initiate acute lymphoblastic leukemia. *J Exp Med* 208(6):1135–1149
36. Feinberg AP, Tycko B (2004) The history of cancer epigenetics. *Nat Rev Cancer* 4(2):143–153
37. Schneider P et al (2011) What role for angiogenesis in childhood acute lymphoblastic leukaemia? *Adv Hematol* 2011:274628
38. Sanda T et al (2013) TYK2-STAT1-BCL2 pathway dependence in T-cell acute lymphoblastic leukemia. *Cancer Discov* 3(5):564–577
39. Jabagi MJ et al (2019) Risk of hematologic malignant neoplasms after postoperative treatment of breast cancer. *Cancers (Basel)* 11(10)
40. Smith MA, McCaffrey RP, Karp JE (1996) The secondary leukemias: challenges and research directions. *J Natl Cancer Inst* 88(7):407–418
41. Croce CM (2008) Oncogenes and cancer. *N Engl J Med* 358(5):502–511
42. Calin GA et al (2002) Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A* 99(24):15524–15529
43. Akbari Moqadam F et al (2013) MiR-125b, miR-100 and miR-99a co-regulate vincristine resistance in childhood acute lymphoblastic leukemia. *Leuk Res* 37(10):1315–1321
44. Benetatos L, Vartholomatos G (2013) MicroRNAs mark in the MLL-rearranged leukemia. *Ann Hematol* 92(11):1439–1450
45. Dou L et al (2013) MicroRNA-205 downregulates mixed-lineage-AF4 oncogene expression in acute lymphoblastic leukemia. *Onco Targets Ther* 6:1153–1160
46. Li X et al (2013) Overexpression of miR-708 and its targets in the childhood common precursor B-cell ALL. *Pediatr Blood Cancer* 60(12):2060–2067
47. Han S et al (2010) Polymorphisms in innate immunity genes and risk of childhood leukemia. *Hum Immunol* 71(7):727–730
48. Strachan DP (1989) Hay fever, hygiene, and household size. *BMJ* 299(6710):1259–1260
49. Kinlen LJ (1995) Epidemiological evidence for an infective basis in childhood leukaemia. *Br J Cancer* 71(1):1–5
50. Greaves M (2006) Infection, immune responses and the aetiology of childhood leukaemia. *Nat Rev Cancer* 6(3):193–203
51. Adelman AS et al (2005) Urbanisation and incidence of acute lymphocytic leukaemia among United States children aged 0–4. *Br J Cancer* 92(11):2084–2088
52. Kroll ME et al (2006) Childhood leukemia incidence in Britain, 1974–2000: time trends and possible relation to influenza epidemics. *J Natl Cancer Inst* 98(6):417–420
53. Urayama KY et al (2011) Early life exposure to infections and risk of childhood acute lymphoblastic leukemia. *Int J Cancer* 128(7):1632–1643
54. Chang JS et al (2012) Allergy and risk of childhood acute lymphoblastic leukemia: a population-based and record-based study. *Am J Epidemiol* 176(11):970–978
55. Boothe VL et al (2014) Residential traffic exposure and childhood leukemia: a systematic review and meta-analysis. *Am J Prev Med* 46(4):413–422
56. Pedersen C et al (2015) Residential exposure to extremely low-frequency magnetic fields and risk of childhood leukaemia, CNS tumour and lymphoma in Denmark. *Br J Cancer* 113(9):1370–1374
57. Kharazmi E et al (2012) Familial risks for childhood acute lymphocytic leukaemia in Sweden and Finland: far exceeding the effects of known germline variants. *Br J Haematol* 159(5):585–588
58. Hunger SP, Mullighan CG (2015) Acute lymphoblastic leukemia in children. *N Engl J Med* 373(16):1541–1552

59. Buffler PA et al (2005) Environmental and genetic risk factors for childhood leukemia: appraising the evidence. *Cancer Investig* 23(1):60–75
60. Trevino LR et al (2009) Germline genomic variants associated with childhood acute lymphoblastic leukemia. *Nat Genet* 41(9):1001–1005
61. Papaemmanuil E et al (2009) Loci on 7p12.2, 10q21.2 and 14q11.2 are associated with risk of childhood acute lymphoblastic leukemia. *Nat Genet* 41(9):1006–1010
62. Sherborne AL et al (2010) Variation in CDKN2A at 9p21.3 influences childhood acute lymphoblastic leukemia risk. *Nat Genet* 42(6):492–494
63. Xu H et al (2013) Novel susceptibility variants at 10p12.31-12.2 for childhood acute lymphoblastic leukemia in ethnically diverse populations. *J Natl Cancer Inst* 105(10):733–742
64. Rudant J et al (2015) ARID5B, IKZF1 and non-genetic factors in the etiology of childhood acute lymphoblastic leukemia: the ESCALE study. *PLoS One* 10(3):e0121348
65. Jin MW et al (2016) A review of risk factors for childhood leukemia. *Eur Rev Med Pharmacol Sci* 20(18):3760–3764
66. INCA, Estimativa 2018: incidência de câncer no Brasil. 2018
67. Chokkalingam AP et al (2011) Haplotypes of DNA repair and cell cycle control genes, X-ray exposure, and risk of childhood acute lymphoblastic leukemia. *Cancer Causes Control* 22(12):1721–1730
68. Chang JS et al (2010) Genetic polymorphisms in adaptive immunity genes and childhood acute lymphoblastic leukemia. *Cancer Epidemiol Biomark Prev* 19(9):2152–2163
69. Wiemels J (2012) Perspectives on the causes of childhood leukemia. *Chem Biol Interact* 196(3):59–67
70. Heck JE et al (2014) Risk of leukemia in relation to exposure to ambient air toxics in pregnancy and early childhood. *Int J Hyg Environ Health* 217(6):662–668
71. Sandler DP, Ross JA (1997) Epidemiology of acute leukemia in children and adults. *Semin Oncol* 24(1):3–16
72. Infante-Rivard C et al (1999) Risk of childhood leukemia associated with exposure to pesticides and with gene polymorphisms. *Epidemiology* 10(5):481–487
73. Hashibe M et al (2005) Epidemiologic review of marijuana use and cancer risk. *Alcohol* 35(3):265–275
74. Metayer C et al (2013) Tobacco smoke exposure and the risk of childhood acute lymphoblastic and myeloid leukemias by cytogenetic subtype. *Cancer Epidemiol Biomark Prev* 22(9):1600–1611
75. Chunxia D et al (2019) Tobacco smoke exposure and the risk of childhood acute lymphoblastic leukemia and acute myeloid leukemia: a meta-analysis. *Medicine (Baltimore)* 98(28):e16454
76. Koo HH (2011) Philadelphia chromosome-positive acute lymphoblastic leukemia in childhood. *Korean J Pediatr* 54(3):106–110
77. Quiroz E et al (2019) The emerging story of acute lymphoblastic leukemia among the Latin American population - biological and clinical implications. *Blood Rev* 33:98–105
78. Greaves MF, Alexander FE (1993) An infectious etiology for common acute lymphoblastic leukemia in childhood? *Leukemia* 7(3):349–360
79. Ling C et al (2019) Residential mobility in early childhood and the impact on misclassification in pesticide exposures. *Environ Res* 173:212–220
80. Park AS et al (2020) Prenatal pesticide exposure and childhood leukemia - a California statewide case-control study. *Int J Hyg Environ Health* 226:113486
81. Han FF et al (2013) Effects of the NQO1 609C>T polymorphism on leukemia susceptibility: evidence from a meta-analysis. *Asian Pac J Cancer Prev* 14(9):5311–5316
82. Petridou ET et al (2018) Advanced parental age as risk factor for childhood acute lymphoblastic leukemia: results from studies of the childhood leukemia international consortium. *Eur J Epidemiol* 33(10):965–976
83. Orsi L, Magnani C, Petridou ET, Dockerty JD, Metayer C, Milne E, Bailey HD, Dessypris N, Kang AY, Wesseling C, Infante-Rivard C, Wunsch-Filho V, Mora AM, Spector LG, Clavel J (2018) Living on a farm, contact with farm animals and pets, and childhood acute lymphoblastic leukemia: pooled and meta-analyses from the Childhood Leukemia International Consortium. *Cancer Med* 7(6):2665–2681. <https://doi.org/10.1002/cam4.1466>

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